




ORIGINAL ARTICLE

The Effect of β -Cryptoxanthin on Testicular Ischemia-Reperfusion Injury in a Rat Model: Evidence from Testicular Histology

Kianoush Mohammadnejad¹, Rahim Mohammadi ¹, Ali Soleimanzadeh², Ali Shalizar-Jalali³, Farshid Sarrafzadeh-Rezaei¹

¹ Department of Surgery and Diagnostic Imaging, Faculty of Veterinary Medicine, Urmia University, Urmia, Iran. ² Department of Theriogenology, Faculty of Veterinary Medicine, Urmia University, Urmia, Iran. ³ Department of Basic Sciences, Faculty of Veterinary Medicine, Urmia University, Urmia, Iran.

ARTICLE INFO

ABSTRACT

Article History:

Received: 12 May 2024
Revised: 29 July 2024
Accepted: 12 August 2024

Keywords:

β -cryptoxanthin
Ischemia-reperfusion
Torsion
Testis

Testicular torsion and detorsion are significant clinical issues for infertile man. Torsion of the spermatic cord is an emergency condition resulting from rotation of the testis and epididymis around the axis of the spermatic cord. A rat testis model was used to assess effects of β -cryptoxanthin on ischemia-reperfusion injury. Twenty healthy male Wistar rats were included and randomized into four investigational groups ($n = 5$): Group SHAM: In this group, midline incision of the scrotum was performed and the testicles were taken out for 2 hours with a 720-degree rotation. Group ISCHEMIA: In this group, midline incision of the scrotum was performed and the testicles were taken out and undergone ischemia for 2 hours with a 720-degree rotation. Group IS/REP/Oil: In this group, a midline scrotum cut was performed and the testicles were taken out and ischemia was created for 2 hours with a 720-degree rotation and at the end of ischemia 100 μ l of corn oil (β -cryptoxanthin solvent) was injected intraperitoneally. Group IS/REP/CRPTXNTN 2.5: The same as group IS/REP/Oil as well as intraperitoneal administration of 100 μ l of β -cryptoxanthin (2.5 μ g/kg) at the end of ischemia. In all groups, the testes were returned back to scrotum and after 60 days were dissected out and removed for histopathological analyses. β -cryptoxanthin at the dose of 2.5 μ g/kg significantly improved histologic indices compared to other treatment groups ($p < 0.05$). β -cryptoxanthin could be helpful in minimizing ischemia-reperfusion injury in testicular tissue exposed to ischemia.

Introduction

In infertile man, testicular torsion and detorsion are significant clinical issues and torsion of the spermatic cord is an emergency condition resulting from rotation of the testis and epididymis around the axis of the spermatic cord. Up to half of all cases of infertility is due to male factor infertility that in the general population affects one man in 20.¹ The annual incidence of testicular torsion has been reported to be one per 4,000 males and one per 158 males younger than 25 years in which incidence peaks in neonates and adolescents arriving puberty.^{2,3} Immediate operational involvements are compulsory to maintain the blood

flow and avoid the continuous injury on the testis that could result in diminished spermatogenesis in most of cases, hence, everlastingly take down fertility rates.⁴

Accumulation of the stimulated neutrophils that produce reactive oxygen species is a proposed pathogenesis of tissue injury in the course of reperfusion.⁵ The most deleterious result of free radicals, that leads to drop in the membrane potential and subsequently cell injury, is lipid peroxidation in the cell. One of the end products of lipid peroxidation, malondialdehyde (MDA), induces serious cell damage via initiation of polymerization and cross linking in components of membrane.⁶ Free oxygen radicals

 Corresponding author. Email: r.mohammadi@urmia.ac.ir

© Iranian Veterinary Surgery Association, 2025

<https://doi.org/10.30500/ivsa.2024.457134.1405>



This work is licensed under the Creative Commons Attribution-NonCommercial 4.0 International License. To view a copy of this license, visit <http://creativecommons.org/licenses/by-nc/4.0/>

react with DNA and form 8-hydroxyguanine (8-OHGua) that is one of the injurious products of DNA.⁷ Despite continuous production of free oxygen radicals in cells, the existence of endogenous antioxidant defense systems help preserve tissues from the detrimental consequences of the free oxygen radicals.⁸

Usage of potent antioxidants has been reported by others. Protective effects of rosmarinic acid on testicular torsion-detorsion has been demonstrated.⁹ It has been reported that L-carnitine had beneficial effects on ischemia-reperfusion injuries following testicular torsion in a rat model.¹⁰ It has been shown that betaine could protect testicular tissue damage induced by testicular torsion in rat.¹¹

β -Cryptoxanthin is an oxygenated carotenoid with a chemical structure similar to, but more polar than, β -carotene. Although β -carotene is present in large amounts in numerous fruits and vegetables, β -cryptoxanthin is found at high concentrations in only a small number of foods. β -Cryptoxanthin can accept energy from singlet oxygen. The evidence that β -cryptoxanthin is an antioxidant in vitro at most physiological concentrations is persuasive. However, a recent review suggested that there is a difference between nonvitamin A-forming carotenoids (such as lycopene) and vitamin A-forming carotenoids (such as β -cryptoxanthin). Nonvitamin A-forming carotenoids protected against DNA damage in almost all circumstances, but vitamin A-forming carotenoids protected against DNA damage in some cases and enhanced DNA damage in others, especially when they were present at high concentrations.^{12,13}

To the best knowledge of authors, the literature is poor regarding intraperitoneal administration of β -cryptoxanthin on testicular ischemia/reperfusion injury. Therefore, the present study was designed to determine whether β -cryptoxanthin could in fact help protect ischemia/reperfusion induced testicular damage in an animal model.

Materials and Methods

Ethical Considerations

The procedures were approved by the Institutional Animal Care and Use Committee of the University under code number# IR-UU-AEC-3/13 dated 14/-5/2024. All procedures were performed under conditions to minimize any potential suffering of the animals. Ketamine (500 mg/kg; IP) and xylazine (50.00 mg/kg; IP) were administered to euthanize the animals.¹⁴

Design of Study, Randomization and Grouping of Animals

An ambient temperature of (23 \pm 3) °C, constant air humidity and a natural day/night cycle were provided for

two weeks prior and within the experiments and the animals were kept in individual plastic cages with free access to standard rodent laboratory food and tap water. All assessments were conducted by blinded observers unaware of the analyzed groups. Twenty healthy male Wistar rats were included and randomized into four investigational groups (n = 5): Group SHAM: In this group, midline incision of the scrotum was performed and the testicles were taken out for 2 hours with a 720-degree count clock wise rotation. Group ISCHEMIA: In this group, midline incision of the scrotum was performed and the testicles were taken out and undergone ischemia for 2 hours with a 720-degree count clock wise rotation. Group IS/REP/Oil: In this group, a midline scrotum cut was performed and the testicles were taken out and ischemia was created for 2 hours with a 720-degree rotation and at the end of ischemia 100 μ l of corn oil (β -cryptoxanthin solvent) was injected intraperitoneally. Group IS/REP/CRPTXNTN 2.5: The same as group IS/REP/Oil as well as intraperitoneal administration of 100 μ l of β -cryptoxanthin (2.5 μ g/kg) at the end of ischemia. In all groups, the testes were returned back to scrotum and after 60 days the left testis was dissected out and removed for histopathological analyses and right testis was dissected out and removed for sperm parameters evaluations.

Surgical Procedures

Animals were anesthetized by intraperitoneal administration of ketamine-xylazine (ketamine 5%, 90 mg/kg and xylazine 2%, 5 mg/kg). The procedure was carried out based on the guidelines of the Ethics Committee of the International Association for the Study of Pain.¹⁵ The ethical Committee of the University approved all the experiments.

The testis was exteriorized through scrotum, the gubernaculum was divided and the testis was freed from the epididymo-testicular membrane. The testes were subjected to 720° torsion and maintained wet by a gauze soaked with sterile normal saline. At the suitable time the testes were rotated back to the natural position for reperfusion. One testis was removed 60 days post operation and evaluations were based on analyses of histopathologic studies.

Histological Assessments

Following 60 days reperfusion and euthanasia of the animals, 10% buffered formalin was used to fix the testes for 24 hours. The tissue samples were then processed, embedded in paraffin and a 5- μ m semi-thin sections were prepared. The samples were then stained routinely with hematoxylin-eosin and Masson's trichrome. The sections were then observed under a light microscope and the following parameters were assessed: Testis weight,

epididymis weight, Testis/body weight, Johnsen score, Cosentino score, seminiferous tubule diameter, Sertoli cell index, repopulation index, mitotic index, Leydig cell nuclear, tubular differentiation index, spermiogenesis index.

Statistical Analysis

Data were analyzed by a commercially available Statistical Package for Social Sciences (SPSS Inc., Chicago, IL, USA) program for Windows software. p -values < 0.05 were regarded as statistically significant. One-way analysis of variance (ANOVA) test was performed and *post hoc* multiple comparisons were done with least-squares differences.

Results

The impact of ischemia/reperfusion on spermatogenesis in testicular tissue is shown in Figure 1. The level of damage caused by testicular torsion/detorsion compared to the SHAM and testicular biopsy scores for the testicular sections from experimental groups are also presented in Table 1. Cross sections from testes of ISCHEMIA group showed seminiferous tubules with severe sloughing and disorganization. Damage was at the spermatozoa, spermatids and some spermatocytes level. The interstitial was disorganized and contained some blood. In, seminiferous tubules showed normal spermatogenesis, exhibiting all stages of spermatogenic cells including

abundant spermatozoa. Some sloughing and tissue disorganization were observed ($p < 0.05$). Testis weight, epididymis weight, testis/body weight, Johnsen score, Cosentino score, seminiferous tubule diameter, Sertoli cell index, repopulation index, mitotic index, Leydig cell nuclear, tubular differentiation index, and spermiogenesis index in IS/REP/CRPTXNTN 2.5 group showed significant improvements compared to other groups ($p < 0.05$).

Discussion

In the present study, it was investigated whether intraperitoneal administration of various concentrations of β -cryptoxanthin was useful in the prevention of testicular damage in ischemia/reperfusion conditions in rat testes and it was found that the concentration of 2.5 $\mu\text{g}/\text{kg}$ showed the most beneficial effects. Histopathologic assessments were performed in experimental groups. The findings for IS/REP/CRPTXNTN 2.5 group were significantly different from those of other groups showing that β -cryptoxanthin at the concentration of 2.5 $\mu\text{g}/\text{kg}$ could improve damages induced by ischemia.

Testicular torsion is a urological emergency that induces biochemical and morphological changes.¹⁶ Testicular torsions can affect males of any age, however, it occurs more often in neonates, boys and young men.¹⁷ The best of our knowledge the impact on prognosis of age at testicular torsion is unknown. The prognosis of testicular torsion is related to the duration and degree of

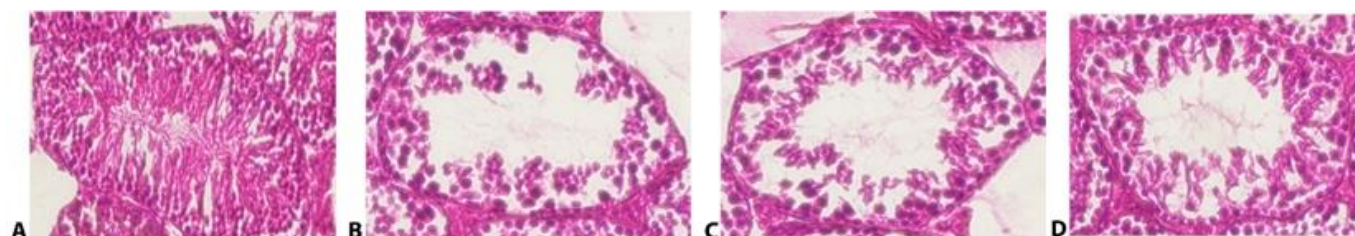


Figure 1. Representative histologic micrograph in different experimental groups. A) SHAM, B) ISCHEMIA group; C) IS/REP/Oil group and D) IS/REP/CRPTXNTN 2.5 group (hematoxylin and eosin staining, 400 \times).

Table 1. Histological parameters. Values are expressed as mean \pm SEM.

Parameters	SHAM	ISCHEMIA	IS/REP/Oil	IS/REP/CRPTXNTN 2.5
Testis weight (g)	1.529 \pm 0.06 ^a	0.507 \pm 0.04 ^c	0.504 \pm 0.06 ^c	0.593 \pm 0.02 ^d
Epididymis weight (g)	0.783 \pm 0.04 ^a	0.261 \pm 0.05 ^d	0.267 \pm 0.02 ^d	0.324 \pm 0.04 ^d
Testis/body weight (%)	0.38 \pm 0.21 ^a	0.20 \pm 0.11 ^d	0.20 \pm 0.13 ^d	0.23 \pm 0.16 ^d
Johnsen score	8.39 \pm 0.19 ^a	3.88 \pm 0.36 ^d	3.95 \pm 0.29 ^d	4.65 \pm 0.32 ^d
Cosentino score	1.07 \pm 0.03 ^d	3.83 \pm 0.19 ^a	3.96 \pm 0.31 ^a	3.25 \pm 0.24 ^b
Seminiferous tubule diameter (STsD) (μm)	254.41 \pm 13.49 ^a	114.49 \pm 17.61 ^d	115.83 \pm 13.46 ^d	130.19 \pm 16.02 ^d
Sertoli cell index (SCI)	25.31 \pm 1.36 ^a	3.01 \pm 0.07 ^e	2.84 \pm 0.09 ^e	4.35 \pm 0.25 ^d
Repopulation index (RI)	79.21 \pm 2.18 ^a	35.93 \pm 1.44 ^d	36.89 \pm 1.61 ^d	37.31 \pm 1.78 ^d
Mitotic index (MI)	3.80 \pm 0.43 ^a	0.79 \pm 0.25 ^d	0.83 \pm 0.49 ^d	1.05 \pm 0.04 ^d
Leydig cell nuclear (LCND) (cell/ mm^2)	9.46 \pm 0.31 ^a	2.88 \pm 0.15 ^d	2.85 \pm 0.46 ^d	3.19 \pm 0.63 ^d
Tubular differentiation index (TDI) (%)	80.05 \pm 2.69 ^a	45.78 \pm 1.63 ^e	47.50 \pm 1.39 ^e	51.62 \pm 1.73 ^d
Spermiogenesis index (SPI) (%)	75.13 \pm 3.84 ^a	45.21 \pm 1.40 ^{de}	44.04 \pm 1.60 ^e	48.62 \pm 1.45 ^d

^{a-e} Different superscripts within the same row demonstrate significant differences ($p < 0.05$).

torsion, resulting in different levels of parenchymal injury by oxidative stress.¹⁸ Therefore, beyond rapid diagnosis and treatment several methods have been developed to minimize the injury caused by testicular torsion.^{19,20} Rat testes differ somewhat from human testes, rats have been widely used as experimental models in testicular torsion studies because lesions in rat testes are comparable to those in human testes after torsion.²¹

Several antioxidants have been investigated with promising results in rats subjected to testicular torsion.²² It has been demonstrated that blood flow following ischemia starts further damage to the reperfused tissue.²³ Ischemia-reperfusion ends up testicular tissue injury and disturbs sperm quality due to overproduction of reactive oxygen species, neutrophil aggregation, membrane lipid peroxidation, apoptosis, and hypoxia.^{24,25} Additionally, ischemia-reperfusion activates an disproportion between the oxygen supply and demand in mitochondria because of buildup of superoxide in vulnerable organs, aggregation of mitochondrial reactive oxygen species. This functional flaw changes permeability of the cell membrane and upsets cell integrity.²⁶

Two separate phases of reactive oxygen species build up have been proposed in testicular torsion/detorsion. In the first phase, a brief period and correlated with reperfusion of testicular tissue, oxidative stress takes place. However, cellular damages may be reversible. Once the oxidative stress lasts for a prolonged time, several days, the second phase is triggered. In the latter phase, injury to testicular tissue becomes more extensive and irreversible. The findings of the present study were based on the first phase in which reperfusion took place 3 hours following initiation of ischemia.²⁷⁻²⁹

Testicular I/R injury can significantly deteriorate spermatogenesis and may cause infertility. Spermatogenesis was evaluated by the Johnsen tubular score and seminiferous tubule diameter measurements in all experimental groups and found to be significantly damaged in the IS/REP group. However, the Johnsen tubular score and seminiferous tubule diameter were significantly higher in IS/REP/CRPTXNTN 2.5 group in comparison with other treatment groups. This finding indicated that β -cryptoxanthin (2.5 $\mu\text{g}/\text{kg}$) administration before detorsion decreased spermatogenesis damage and also could prevent testicular I/R injury induced infertility.

Changes in histological parameters (Testis weight, epididymis weight, Testis/body weight, Johnsen score, Cosentino score, seminiferous tubule diameter, Sertoli cell index, repopulation index, meiotic index, Leydig cell nuclear, tubular differentiation index, spermiogenesis index) have been used in the evaluation of the cell condition.³⁰ In consistent with the findings of the present study, it was reported that β -cryptoxanthin alleviated

myocardial ischemia/reperfusion in rats.³¹ In a rat model of hepatic I/R injury, supplementation with β -carotene has been shown to increase the level of antioxidants.³² Moreover, β -carotene was demonstrated to protect the protects the gastric mucosa against ischemia-reperfusion injury in rats by inhibiting the peroxidation of lipids, improving the activity of antioxidant enzymes and preventing the infiltration of neutrophils.³³ Taken together, the findings suggested that β -carotene had multiple beneficial roles as a gastro protective agent, and might be responsible for the reduction in pro-inflammatory cytokine release and the increase in anti-inflammatory cytokines.³⁴

Substances are administered by a wide variety of routes. A key factor determining the route selected is whether the agent is being administered for a local or systemic (either enteral or parenteral effect. Parenteral administration methods typically produce the highest bioavailability of substances because these methods avoid the first-pass effect of hepatic metabolism, which occurs commonly with orally administered chemicals and therapeutics. Intraperitoneal administration seems more effective and available where oral administration of an agent may cause difficulties. It is clear that transperitoneal absorption of the agent is far faster than oral administration.²² It seems time saving is very important in emergency conditions like ovarian torsion.

Although in the present study the outcomes were promising, however, extensive molecular assessments are required to evaluate outcomes of intraperitoneal administration of β -cryptoxanthin on testicular ischemia/reperfusion injury that remained unknown. These could be regarded as limitations of our study.

In conclusion, findings obtained from all the experimental groups indicated that intraperitoneal administration of β -cryptoxanthin at concentration of 2.5 $\mu\text{g}/\text{kg}$ could be helpful in minimizing ischemia-reperfusion injury in testicular tissue exposed to ischemia. Some works were completed in the present study, however, the exact underlying mechanism of β -cryptoxanthin on improving testicular function might be more complicated than our findings.

Acknowledgments

This work was fully supported by Vice-Chancellor for Research and Technology of Islamic Azad University, Urmia branch that is acknowledged by the authors.

Conflict of Interest

None.

References

1. McLachlan RI, de Krester DM. Male infertility: the case for continued research. *Medical Journal of Australia*. 2001;

- 174(3): 116-117. doi: 10.5694/j.1326-5377.2001.tb143180.x
2. Srinath H. Acute scrotal pain. *Australian Family Physician*. 2013; 42(11): 790-792.
 3. Arap MA, Vicentini FC, Cocuzza M, Hallak J, Athayde K, Lucon AM, Arap S, Srougi M. Late hormonal levels, semen parameters, and presence of antisperm antibodies in patients treated for testicular torsion. *Journal of Andrology*. 2007; 28(4): 528-532. doi: 10.5534/wjmh.190037
 4. Parhizkar P, Mohammadi R, Shahrooz R, Mohammadi V. Effects of pyrroloquinoline quinone (PQQ) on ischemia-reperfusion injury in rat ovaries: histological and biochemical assessments. *Bulletin of Emergency and Trauma*. 2019; 7(1): 35-40. doi: 10.29252/beat-070105
 5. Vakili-Sadeghi E, Najafpour A, Mohammadi R. Protective effects of propolis on ischemia-reperfusion injury in a rat testicular torsion and detorsion model. *Veterinary Research Forum*. 2023; 14(7): 389-395. doi: 10.30466/vrf.2023.1972630.3682
 6. Girotti AW. Lipid hydroperoxide generation, turnover, and effector action in biological systems. *Journal of Lipid Research*. 1998; 39(8): 1529-1542.
 7. Huang HY, Helzlsouer KJ, Appel LJ. The effects of vitamin C and vitamin E on oxidative DNA damage: results from a randomized controlled trial. *Cancer Epidemiology, Biomarkers and Prevention*. 2000; 9(7): 647-652.
 8. Ames BN, Shigenaga MK, Hagen TM. Oxidants, antioxidants, and the degenerative diseases of aging. *Proceedings of the National Academy of Sciences of the USA*. 1993; 90(17): 7915-7922. doi: 10.1073/pnas.90.17.7915
 9. Raisi A, Davoodi F, Mohammadi R. Protective effects of rosmarinic acid on testicular torsion-detorsion in an animal model. *Iranian Journal of Veterinary Surgery*. 2022; 17(2): 80-86. doi: 10.30500/IVSA.2022.327175.1293
 10. Kazemi-Darabadi S, Asadpour R, Shahbazfar AA, Alizadeh S. Effects of L-carnitine and betamethasone on ischemia-reperfusion injuries and sperm parameters following testicular torsion in a rat model. *Veterinary Research Forum*. 2019; 10(2): 125-132. doi: 10.30466/vrf.2019.74767.2002
 11. Sarlak M, Roumiani E, Kheradmand A, Dezfoulan O, Alirezaei M, Raisi A, Davoodi, F. Evaluating the effects of betaine on testicular ischemia/reperfusion injury induced by torsion/detorsion in the rat. *Andrologia*. 2022; 54(10): e14559. doi: 10.1111/and.14559
 12. Burri B J, La Frano MR, Zhu C. Absorption, metabolism, and functions of β -cryptoxanthin. *Nutrition Reviews*. 2016; 74(2), 69-82. doi: 10.1093/nutrit/nuv064
 13. Azqueta A, Collins AR. Carotenoids and DNA damage. *Mutation Research*. 2012; 733(1-2): 4-13. doi: 10.1016/j.mrfmmm.2012.03.005
 14. Animal euthanasia policy. Approved and adopted. [Accessed May 03, 2023]. Available at: <https://research.wustl.edu/wp-content/uploads/2017/05/Euthanasia.pdf>.
 15. Ahmad MB, Tay MY, Shameli K, Hussein MZ, Lim JJ. Green synthesis and characterization of silver/chitosan/polyethylene glycol nanocomposites without any reducing agent. *International Journal of Molecular Sciences*. 2011; 12(8): 4872-4884. doi: 10.3390/ijms12084872
 16. Cosentino MJ, Nishida M, Rabinowitz R, Cockett AT. Histopathology of prepubertal rat testes subjected to various durations of spermatic cord torsion. *Journal of Andrology*. 1986; 7(1): 23-31. doi: 10.1002/j.1939-4640.1986.tb00862.x
 17. Cummings JM, Boullier JA, Sekhon D, Bose K. Adult testicular torsion. *Journal of Urology*. 2002; 167(5): 2109-2110.
 18. Aktaş BK, Bulut Ş, Bulut S, Baykam MM, Özden C, Şenes M, Yücel D, Memiş A. The effects of N-acetylcysteine on testicular damage in experimental testicular ischemia/reperfusion injury. *Pediatric Surgery International*. 2010; 26(3): 293-298. doi: 10.1007/s00383-009-2538-0
 19. Lee JW, Kim JI, Lee YA, Lee DH, Song CS, Cho YJ, Han JS. Inhaled hydrogen gas therapy for prevention of testicular ischemia/reperfusion injury in rats. *Journal of Pediatric Surgery*. 2012; 47(4): 736-742. doi: 10.1016/j.jpedsurg.2011.09.035
 20. Barlas M, Hatiboglu C. The effect of nitric oxide in testicular ischemia-reperfusion injury. *International Urology and Nephrology*. 2002; 34(1): 81-86. doi: 10.1023/a:1021311029572
 21. Wang WW, Qiao SY, Li DF. Amino acids and gut function. *Amino Acids*. 2009; 37(1): 105-110. doi: 10.1007/s00726-008-0152-4
 22. Vitaglione P, Ottanelli B, Milani S, Morisco F, Caporaso N, Fogliano V. Dietary trans-resveratrol bioavailability and effect on CCl4-induced liver lipid peroxidation. *Journal of Gastroenterology and Hepatology*. 2009; 24(4): 618-622. doi: 10.1111/j.1440-1746.2008.05598.x
 23. Ünsal A, Eroglu M, Avci A, Cimentepe E, Guven C, Derya Balbay M, Durak I. Protective role of natural antioxidant supplementation on testicular tissue after testicular torsion and detorsion. *Scandinavian Journal of Urology and Nephrology*. 2006; 40(1): 17-22. doi: 10.1080/00365590500407514
 24. Cuzzocrea S, Riley DP, Caputi AP, Salvemini, D. Antioxidant therapy: a new pharmacological approach in shock, inflammation, and ischemia/reperfusion injury. *Pharmacological Reviews*. 2001; 53(1): 135-159.
 25. Turner TT, Bang HJ, Lysiak JL. The molecular pathology of experimental testicular torsion suggests adjunct therapy to surgical repair. *Journal of Urology*. 2004; 172(6 Part 2): 2574-2578. doi: 10.1097/01.ju.0000144203.30718.19
 26. Jung JE, Kim GS, Chen H, Maier CM, Narasimhan P, Song YS, Song YS, Niizuma K, Katsu M, Okami N, Yoshioka H, Sakata H, Goeders CE, Chan PH. Reperfusion and neurovascular dysfunction in stroke: from basic mechanisms to potential strategies for neuroprotection. *Molecular Neurobiology*. 2010; 41(2-3): 172-179. doi: 10.1007/s12035-010-8102-z
 27. Abdel-Gaber SA, Mohammed RK, Refaie MM. Mechanism mediating the protective effect of diacerein in ischemia-reperfusion-induced testicular injury in rats. *Life Sciences*. 2018; 209: 57-62. doi: 10.1016/j.lfs.2018.07.060
 28. Cutrì JC, Boveris A, Zingaro B, Corvetti G, Poli G. *In situ* determination by surface chemiluminescence of temporal relationships between evolving warm ischemia-reperfusion injury in rat liver and phagocyte activation and recruitment. *Hepatology*. 2000; 31(3): 622-632. doi: 10.1002/hep.510310312
 29. Kim YH, Kim GH, Shin JH, Kim KS, Lim JS. Effect of Korean red ginseng on testicular tissue injury after torsion and detorsion. *Korean Journal of Urology*. 2010; 51(11): 794-799. doi: 10.4111/kju.2010.51.11.794
 30. Celik O, Turkoz Y, Hascalik S, Hascalik M, Cigremis Y, Mizrak B, Yologlu S. The protective effect of caffeic acid phenethyl ester on ischemia-reperfusion injury in rat ovary. *European Journal of Obstetrics, Gynecology, and Reproductive Biology*. 2004; 117(2): 183-188. doi: 10.1016/j.ejogrb.2004.05.007
 31. Zhang F, Shi D, Wang X, Zhang Y, Duan W, Li Y. β -cryptoxanthin alleviates myocardial ischaemia/reperfusion injury by inhibiting NF- κ B-mediated inflammatory signalling in rats. *Archives of Physiology and Biochemistry*, 2022; 128(4): 1128-1135. doi:

- 10.1080/13813455.2020.1760302
32. Codoñer-Franch P, Muñoz P, Gasco E, Domingo JV, Valls-Belles V. Effect of a diet supplemented with alpha-tocopherol and beta-carotene on ATP and antioxidant levels after hepatic ischemia-reperfusion. *Journal of Clinical Biochemistry and Nutrition*. 2008; 43: 13-18. doi: 10.3164/jcbn.2008038
33. Vardi N, Parlakpınar H, Öztürk F, Ates B, Gül M, Çetin A, Erdoğan A, Ötlu A. Potent protective effect of apricot and beta-carotene on methotrexate-induced intestinal oxidative damage in rats. *Food and Chemical Toxicology*. 2008; 46: 3015-3022. doi: 10.1016/j.fct.2008.05.039
34. Seyyed Ali M, Neisi N, Darbor M, Hassanpour M, Makvandi M, Solgi G. beta-carotene protects the gastric mucosa against ischemia-reperfusion injury in rats. *European Cytokine Network*. 2012; 23(4): 173-178. doi: 10.1684/ecn.2012.0317